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Mineral Contents Analysis of *Cyperus esulentus* (Tigernut)

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ABSTRACT

The objective of this work was to determine the amount of minerals such as, iron, copper, nickel, chromium, zinc, manganese lead, cadmium, calcium, sodium, potassium, cobalt, magnesium, present in tigernut using Atomic Absorption Spectrophotometer (AAS), while gravimetric method was used to determine phosphorus and chloride. The sample for the analysis was prepared by wet digestion method based on the addition of nitric acid followed by oven heating the temperature 150°C. The standard solutions of the above minerals were equally synthesized. These standards were measured in the AAS machine to strike a balance between absorbance against concentration which was utilized in quantitatively determining the amount in prepared sample of tigernut. The result gave the following minerals as Fe 0.002, Cu 0.02, Ni 0.03, Zn 0.038 Mn 0.004, Cd 0.16, Ca 0.047, Na 0.01, K 0.18, Co 0.02, Mg 0.02, P 32.89, Cl 2.24 all in (mg/g).

Keywords: Minerals, contents, analysis, tigernut.

INTRODUCTION

Tigernut (Cyperus esculentum) is a perennial grass-like plant spheroid tubers, pale yellow cream kernel surrounded by a fibrous sheath. It is also known as yellow nut sedge, earth or ground almonds. [1] reported that chufa came to Spain from Africa. Tigernut is found in the wild and cultivated in Africa. South America, Europe and Asia. It along rivers grows cultivated on a small scale by rural farmers mostly in the northern states of Nigeria. It is locally called "aya" in Hausa; "aki awusa" in Igbo; "ofio" in Yoruba and "isipaccara" in Efik. Tigernuts are edible, sweet, flavoured nutty, tubers which carbohydrate, contain protein, sugars, and lots of oil and fiber [2]. [3] showed that tigernuts have been cultivated for food and drink for men and planted for hogs for many years in Spain and that the lovely milky elixir is served in health Spas, Restaurants Pubs. and refreshing beverage. Unfortunately, despite these potentials in tigernuts it has been a neglected crop in Nigeria. This probably may be due to inadequate knowledge

utilization and nutritional value. It is an important food crop for certain tribes in Africa, often collected and eaten raw, baked as a vegetable, roasted or dried and ground to flour. The ground floor is mixed with sorghum to make porridge, icecream, sherbet or milky drink. It is mostly consumed raw as snack without knowledge of the food and nutritional quality [4]. It has also found to possess therapeutic quality [5]. Moore stated that "the expansion of tigernut milky drinks will significantly help the research linking tigernut milk to healthier cholesterol levels and other non-dairy manufacturers. This could also gain a boost from an increased consumer interest healthy foods".

Variety of food products can be derived from tiger nut tubers though little-documentation there is Various food processing techniques can be applied to tiger processing to modify develop appearance, its natural flavour, stimulate the digestive juices, add variety to the menu. make it easily digestible and bioavailable, destroy harmful microorganisms, improve its nutritional quality and prevent

decomposition. This project work intends to basically evaluate mineral contents and utilization of tigernuts

STATEMENT OF PROBLEMS

Recognition of food crops with good mineral content has been a serious problem to the nation due to little or knowledge on most common crops with this mineral value such as tigernut. Though rich in mineral contents, tigernuts have been an underutilized food crop in Nigeria. It is mostly eaten raw as snack and this food crop has a great potential in improving someone's nutritional status. Adequate nutrition essential for individual development, activity, good health. Nutritionists have demonstrated that major nutritional problems could be solved through exploitation of the nutrition and economic of the local potentials food

resources. Tigernut is one of the under- utilized tubers with great potentials for domestic commercial purposes. There is no documentation of a successful product made from tigernuts in the Nigerian market. Α successful product, offers a benefit that is perceptible and valued by consumer [6]. There is little documentation on the nutritional quality and versatility of tigernuts in food preparation despite its availability. However, tigernut is still one of the least popular tubers in Nigeria and hence the need for this research which intends evaluate the mineral content and utilization of tiger nuts.

OBJECTIVES OF STUDY

The objective of this study is to evaluate the mineral potentials of

Cyperus esculentum (tigernut).

MATERIALS AND METHODS Sample Collection

Dried tigernut used were purchased from Ogbete main Market in Enugu Stale. Nigeria in February 2018 and

Reagent Preparation
Zinc Chloride Solution (100ppm)

Zinc chloride (0.3g) was weighed using OHAUS weighing balance and introduced into 30ml volumetric flask. distilled water (100ml) was measured with a measuring cylinder and introduced the flask, then shaken to dissolve the zinc chloride. solution was made up to mark with distilled water. Portions of solution (2.5ml, 2ml, and 1ml) were separately measured with measuring cylinder and poured into a 100ml volumetric flask. distilled water (70ml) was measured and introduced the flask. It was shaken vigorously and made up to mark with distilled water to obtain series of working standards.

Manganese Solution (100ppm)

Manganese (II) chloride (0.24g) was weighed with OHAUS weighing balance and introduced into a 1000ml volumetric flask, distilled

carried to the laboratory in a polythene bag.

water (100ml) was measured using measuring cylinder and poured into flask. It was shaken to dissolve the manganese (II) chloride. The solution was made up to mark with distilled water. Portions of the solution 0.02ml), $(2.5 \,\mathrm{ml},$ 1.5ml, separately measured with measuring cylinder and poured into a 100ml volumetric flask. distilled (70ml) was measured and introduced into the flask. was shaken It vigorously and made up mark with distilled water to obtain series of working standards.

Magnesium Solution (1000ppm)

Magnesium chloride (0.4g)was OHAUS weighed with weighing balance and introduced into 1000ml volumetric flask, distilled water (100ml) was measured using a measuring cylinder and poured into the flask. It was shaken to dissolve magnesium chloride. solution was made up to mark with distilled water. Portions of

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solution (1.5ml, 1.3ml, 0.8ml), were separately measured with measuring cylinder and poured into a 100ml volumetric flask, distilled water (70ml) was measured and introduced the flask. It was shaken vigorously and made up to mark with distilled water to obtain series of working standards.

Calciuin Solution (100ppm)

Calcium chloride (0.3g) was weighed with OHAUS weighing balance and 1000ml introduced into was volumetric flask, distilled water $(100 \mathrm{ml})$ measured with was measuring cylinder and poured into the flask, then shaken to dissolve the calcium chloride. The solution was made up to ark with distilled water. Portions of the solution (6ml, 3.2ml, and 1.5ml) were separately measured with measuring cylinder and poured into a 100mL volumetric flask, distilled water (70ml) was measured and introduced into the flask. It was shaken vigorously and made up to mark with distilled water obtain series of working standards.

Iron Solution (100ppm)

Iron (II) chloride (0.26g) was weighed with OHAUS weighing balance and introduced into a 1000ml volumetric flask, distilled water (100ml) was measured using with a measuring cylinder and poured into the flask, then shaken to dissolve the iron (II) chloride. The solution was made to mark with distilled water. Portions of the solution (5.2ml, 3ml, and 13ml) separately measured measuring cylinder and poured into a 100mL volumetric flask, distilled water (70ml)was measured introduced into the flask. It was shaken vigorously and made up to mark with distilled water to obtain series of working standards.

Nickel Solution (100ppm)

Nickel (II) chloride (0.22g) was with OHAUS weighing weighed balance and introduced into a 1000ml volumetric flask, distilled water (100ml) was measured with measuring cylinder and poured into the flask, then shaken to dissolve the Nickel (II) chloride. The solution was made up mark with distilled water. Portions of the solution (8ml. 4.2ml. 0.7mlwere separately measured with measuring cylinder and poured into a 100ml volumetric flask, distilled water (70ml) was measured and introduced into the flask. It was shaken vigorously and made up to mark with distilled water obtain series οf working standards.

Chromium Solution (100ppm)

Chromium (III) chloride was weighed with OHAUS weighing balance and introduced into 1000ml volumetric flask, distilled water (100ml) was measured with measuring cylinder and poured into the flask, then shaken to dissolve the chromium (III)chloride. The solution was made up to mark with distilled water. Portions of the solution (5ml, 2.5ml. 0.7ml) were separately measured with measuring cylinder and poured a 100ml volumetric distilled water (70ml) was measured and introduced into the flask. It was shaken vigorously and made up mark with distilled water to obtain series of working standards.

Sodium Solution (100ppm)

Sodium chloride (0.26g) was weighed with (OHAUS) weighing balance and introduced in a 1000ml volumetric flask, distilled water (100ml) was measured with measuring cylinder and poured into flask, then shaken to dissolve, the sodium chloride. The solution was made up to mark with water. Portions of the distilled solution (2ml, 1ml, 0.5ml) were separately measured with measuring cylinder and poured into a 100ml volumetric flask, distilled (70ml) was measured and introduced the flask. It was shaken vigorously and made up to mark with distilled water to obtain series of working standards.

Lead Solution (100ppm)

Lead (II) chloride was weighed with OHAUS weighing balance introduced into a 1000ml volumetric flask, distilled water (100ml) was measured with measuring cylinder and poured into the flask. It was shaken to dissolve the lead (III) chloride. The solution was made up with distilled portions of the solution (21ml, 10ml, 1ml) were separately measured with measuring cylinder and poured into a 100ml volumetric flask, distilled water (70ml) was measured and introduced into the flask. It was shaken vigorously and made up to mark with distilled water to obtain series of working standards,

Copper Solution (ppm)

Copper (II) chloride (0.22g) was OHAUS weighed with weighing balance and introduced into 1000ml volumetric flask, distilled water was measured with measuring cylinder and poured into the flask. It was shaken to dissolve the copper (II) chloride. The solution was made up mark with distilled water. Portions of the solution (10ml. 0.5 ml5.5ml. were separately measured with measuring cylinder and poured into a 100mL volumetric flask, distilled water (70ml) was measured and introduced into the flask. It was shaken vigorously and made up to mark with distilled water obtain series of working standards.

Cadmium Solution (100ppm)

Cadmium chloride (0.16g)was weighed with OHAUS weighing balance and introduced into 1000ml volumetric flask, distilled water (100ml) was measured and poured into the flask, then taken to dissolve the cadmium chloride. The solution was made up to mark with distilled water. Portions of the solution (2ml, $0.1 \mathrm{ml}$ were separately measured with measuring cylinder and poured into a 100ml volumetric flask, distilled water (70ml) was measured and introduced into the flask. It was shaken vigorously and made up to mark with distilled water obtain series of working standards.

Potassiuin Solution (100ppm)

Potassium tetraoxosulphate (VI) (0.45g) was weighed with OHAUS weighing balance and introduced to a 1000ml volumetric flask, distilled water (100ml) was measured with a measuring cylinder and poured into the flask; it was shaken to dissolve the potassium tetraoxosulphate (VI). The solution was made up to make with distilled water. Portions of the solution (5.2ml, 3ml and 1.2ml) were separately measured with measuring

cylinder and poured into a 100ml volumetric flask, distilled water (70ml) was measured and introduced into the flask. It was shaken vigorously and made up to mark with distilled water to obtain series of working standards.

Cobalt Solution

Cobalt (II) chloride (0.22g) was weighed with OHAUS weighing balance and introduced into a 1000ml volumetric flask, distilled water (100ml) was measured with measuring cylinder and poured into the flask. It was shaken to dissolve the cobalt (II) chloride. The solution was made up to mark with distilled water. Portions of the solution (5ml, 0.5 mlwere 3.5ml, separately measured with measuring cylinder and poured into a 100mL volumetric flask, distilled water (70mL) was measured and introduced into the flask. It was shaken vigorously and made up to .sk with distilled water to obtain series of working standards.

Sample Preparation

The sample used for the analysis was crushed in a mortar with pestle to produce homogenized forms of the sample. They were then collected and stored in airtight container until required for use.

Sample Digestion

The sample was grinded into powder using a clean mortar and pestle. 0.5g of the sample was added to 5.0 cm³ of concentrated nitric acid (HNO₃) in a beaker and heated to near dryness at 150°C for six minutes to expel brown gas of NO₂. The resulting solution was allowed to cool, and dissolved a small portion of distilled water and made up to mark in a 100ml volumetric flask.

Analysis of Copper Ion

The copper hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer to give resonance line of wavelength 327nm. The instrument was aspirated with distilled water to clear the aspiration tube of impurities. Three standard solution of different arsenic concentration (10ppm, 5ppm, 0.5ppm) were separately aspirated into the instrument and absorbance against concentration standard curve

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was generated automatically. The aspirated instrument was with distilled water, the sample was then aspirated and absorbance also automatically generated the The copper instrument. concentration of the sample was standard extrapolated from the curve.

Analysis of Zinc Ion

The zinc hollow cathode lamp was setup by selecting arsenic from the library of the atomic absorbance spectrophotometer to give resonance line of wavelength 213.9nm. The instrument aspirated with distilled water to clear the aspiration tube of impurities. Three standard arsenic solution of different concentration (2ppm, lppm, 0.2ppm) were separately aspirated into the instrument and absorbance against concentration standard curve was generated automatically. The instrument was aspirated with distilled water, the sample was then aspirated and absorbance as also generated automatically instrument. The zinc concentration of the sample was extrapolated from the standard curve.

Analysis of Magnesium Ion

The magnesium hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer to give resonance line of wavelength 285.2nm. The instrument was aspirated with distilled water to the aspiration clear tube impurities. Three standard arsenic solution of different concentration (1.5ppm, lppm, 0.5ppm) separately aspirated into the instrument and absorbance against concentration standard curve was automatically. generated instrument was aspirated with distilled water, the sample was then aspirated and absorbance was also generated automatically by The instrument. magnesium concentration of the sample was extrapolated from the standard curve.

Analysis of Cadmium Ion

The cadmium hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer give resonance line of wavelength 228.9nm. The instrument was aspirated with distilled water to clear the aspiration tube οf impurities. Three standard arsenic solution of different concentration (2ppm, lppm, 0.1ppm) separately aspirated into instrument absorbance and against concentration standard curve was generated automatically. instrument was aspirated distilled water, the sample was then aspirated and absorbance was also generated automatically by the instrument. The cadmium of the sample extrapolated from the standard curve.

Analysis of Chromium Ion

The chromium hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer to give line of resonance wavelength 357.9nm. The instrument aspirated with distilled water to clear the aspiration tube of impurities. Three standard arsenic solution of different concentration (5ppm, 2.5ppm, 0.5ppm) separately aspirated into instrument and absorbance against concentration standard curve was automatically. generated The instrument was aspirated distilled water, the sample was then aspirated and absorbance was also generated automatically by the instrument. The chromium concentration of the sample was extrapolated from the standard curve.

Analysis of Calcium Ion

The calcium hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer give to wavelength resonance line οf 427.7nm. The instrument aspirated with distilled water to aspiration clear the tube impurities. Three standard arsenic solution of different concentration (6ppm, 3ppm, lppm) were separately aspirated into the instrument and absorbance against concentration standard curve was generated automatically. The instrument was

aspirated with distilled water, the sample was then aspirated and absorbance .is also generated automatically by the instrument. The calcium concentration of the sample was extrapolated from the standard curve.

Analysis of Iron Ion

The iron hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer give to resonance line of wavelength 372 Ohm. The instrument was aspirated with distilled water to clear the aspiration tube of impurities. Three standard arsenic solution different concentration (5ppm, lppm) 3ppm, were separately aspirated into the instrument and absorbance against concentration standard curve was generated automatically. The instrument was aspirated with distilled water, the aspirated sample was then absorbance as also generated automatically by the instrument. The iron concentration of the sample was extrapolated from the standard curve.

Analysis of Nickel Ion

The nickel hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer to give resonance line of wavelength 341.5nm. The instrument aspirated with distilled water to clear the aspiration tube of impurities. Three standard arsenic solution of different concentration 4pmm, 0.5ppm) were separately aspirated into the instrument and absorbance against concentration curve standard generated was automatically. The instrument was aspirated with distilled water, the sample was then aspirated and was absorbance generated automatically by the instrument. The nickel concentration of the sample was extrapolated from the standard curve.

Analysis of Manganese Ion

The manganese hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer to give a resonance line of wavelength

279.5nm. The instrument aspirated with distilled water clear the aspiration tube impurities. Three standard arsenic solution of different concentration (2.5ppm, 1.5ppm, 0.2ppm) separately aspirated into instrument and absorbance against concentration standard curve was automatically. The generated instrument was aspirated with distilled water, the sample was then aspirated and absorbance was also generated automatically bv The instrument. manganese concentration of the sample was extrapolated from the standard curve.

Analysis of Cobalt Ion

The cobalt hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer to give resonance line of wavelength 322nm. The instrument was aspirated with distilled water to clear the aspiration tube of impurities. Three standard arsenic solution of different concentration (5ppm, 3.5ppm, 0.5pp) were separately aspirated into the instrument and absorbance against concentration standard curve was generated automatically. instrument was aspirated with distilled water, the sample was then aspirated and absorbance was also generated automatically instrument. The cobalt concentration of the sample was extrapolated from the standard curve.

Analysis of Lead Ion

The lead hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer to give of resonance line wavelength 283.3nm. The instrument aspirated with distilled water tο the aspiration clear tuhe οf impurities. Three standard arsenic solution of different concentration (20ppm, l0ppm, lppm) separately aspirated into instrument and absorbance against concentration standard curve was generated automatically. The instrument was aspirated with distilled water, the sample was then aspirated and absorbance was also

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generated automatically by the instrument. The Lead concentration of the sample was extrapolated from the standard curve.

Analysis of Sodium Ion

The sodium hollow cathode lamp was set up by selecting arsenic from the library of the atomic -absorbance spectrophotometer to give of resonance line wavelength The 589.0nm. instrument aspirated with distilled water to clear the aspiration tube οf impurities. Three standard arsenic solution of different concentration 1ppm, 0.5ppm) separately aspirated into instrument and absorbance against concentration standard curve was The generated automatically. instrument was aspirated distilled water, the sample was then aspirated and absorbance was also generated automatically by The sodium instrument. concentration of the sample was extrapolated from the standard curve.

Analysis of Potassium Ion

The potassium hollow cathode lamp was set up by selecting arsenic from the library of the atomic absorbance spectrophotometer give to wavelength resonance line of 769.9nm. The instrument was aspirated with distilled water to the aspiration of tuhe impurities. Three standard arsenic solution of different concentration (5ppm, 3ppm, 1ppm) were separately aspirated into the instrument and absorbance against concentration generated standard curve was automatically. The instrument was aspirated with distilled water, the sample was then aspirated and absorbance was also generated automatically by the instrument. The potassium concentration the sample was extrapolated from the standard curve.

Chloride Determination

The sample weighed was 0.2g and was introduced into a 250ml conical flask and 100ml of distilled water was added. It was heated for one hour (1hr) in a boiling water bath and the extract was filtered. Chloride was determined by titration

with AgNO₃ following [7] 100ml of the water sample was measured into 250ml Erlenmeyer flask. The pH of the water was adjusted to 8.0 using sodium hydroxide. 10ml of 0.1m potassium chromate was added to the solution. The mixture was then titrated with 0.014M Silver trioxonitrate (AgNO₃) until the colour changed from colourless to pinteish yellow as the end point.

Determination of Phosphorus

The sample weighed was 3g using weighing balance and was transferred into a 250ml beaker and 40ml of distilled water was added and stirred to dissolve the sample. The solution was filtered. 45ml of 10% MgSO...H.O solution was added to the filtrate, 150ml of 2m NH3 was added slowly while stirring. The mixture was allowed to settle for 15 minutes at room temperature and the solution was precipitated. Filter paper was weighed and held in a and the solution transferred into filter paper for filtration and 10ml of 75% isopropyl alcohol was used to rinse the beaker and poured into the filter paper. When all the liquid had gone through. the filter paper with the precipitate was removed placed on a marked paper towel while the filtrate was discarded. The filter paper with Magnesium ammonium phosphate hexaphydrate $(MH_{PO}.6H_{0})$ precipitate was thoroughly dried in an oven and weighed with the precipitate tothe weight of the filter paper after heating.

RESULTS

Table 1: The Result of some Minerals contained in Tigernut

Parameter	Values (mg/g)
Fe	0.002
Cu	0.02
Ni	0.03
Cr	0.00
Zn	0.38
Mn	0.004
Pb	0.00
Cd	0.16
Ca	0.47
Na	0.01
K	0.18
Со	0.02
Mg	0.02
P	32.89
Cl	2.24

DISCUSSION

Result in the Table 1 shows the mineral content of tigernut using atomic absorption spectrophotometer (Buck Scientific 210VGP) and gravimetric method of analysis. Result gotten shows that phosphorus (32.89mg/100g) is the most abundant followed by chloride (2.24mg/100g) while chromium and lead were not detected. Calcium (0.47 mg/100 g)and Zinc were mg/100g) the next phosphorus chloride. and (0.002 mg/100g) and Manganese(004 mg/100g) were the least in the result obtained. The result obtained also indicate the presence of other minerals which are the following: Copper (0.02 mg/100g), nickel (0.03 mg/100g)mg/100g), cadmium (0.16 mg/100g),

sodium (0.01 mg/l00g), potassium (0.18 mg/l00g), cobalt (0.02 mg/l00g) and magnesium (0.02 mg/l00g).

Oladele and Aina (2007), reported the mineral element of tigernut flour. Their result showed that sodium had a value of mg/l00g), potassium (216 mg/l00g), calcium (155 mg / l0g) while phosphorus was (121 mg/l00g). Other minerals present are zinc (0.01 mg/l00g), iron (0.65 mg/l00g)and copper (0.02 mg/l00g). These values are higher than what was found in this work except copper which has exact value with what was found in this work and zinc which was lower than what was found in this work.

CONCLUSION

Although tigernut is known to be an underutilized food crop but this study has revealed that tigernut (Cyperus esculentus) contain following minerals: phosphorus, chloride, sodium, calcium, magnesium, potassium, cobalt. copper, cadmium, zinc, manganese,

nickel and iron. From the result of the study, it can be evidently concluded that tigernut is a very good source of some mineral components as mentioned. With these minerals present, it would also serve as useful dietary supplements. www.idosr.org Igwe and Eze-Stephen

REFERENCES

- 1. Addy, E. O. and Eteshola, E., (1984). Nutritive Value of a Mixture of Tigernut Tubers(Cyperusesculenlus L.)and Baobab Seeds(Adansoniadigitata L.). Journal of Science and Agriculture. 35: 437-440.
- 2. Akoma, O., Elekwa, U. A., Afodunrinbi, A. T. and Onyeukwu G.C. (2000) Yoghurt from Coconut and Tiger nut. *Journal of Technology*.**5**:132-134.
- 3. Al-Delaimy, W.K., Rimm, E.B., Willett, W.C., Stampfer, M.J. and Flu. F.B. (2004). Magnesium Intake and Risk of Coronary Ffeart Disease among Men. Journal of the American College of Nutrition.23: 63-70.
- 4. Aletor, V. A. and Ojo, O.I. (2009). Changes in Different Processed Soya Bean(Glycine max) and Loma Beans (Phaseohis lunatus) with Particular reference to Nutritional Constituents. Diet and nutrition.33: 1009 1016.
- 5. Barminas, J.T., Maina, H.M., Tahir, S., Kubmarawa, D. and Tsware, k. (2001). A preliminary investigation into the Biofuel Characteristics of Tigernut (Cyperus esculenius) Oil. Elsevier Bioresource. 79: 87-89.
- Bamgbose, A.M., Eruvbetine, D. and Dada, W. (2003) Utilization of Tiger Nut (Cyperusrotundas, L.) Meal in the Diets for Cockerel Starters. Bioresourse Technology, 89: 245-248.
- 7. Bankole S. A. and Eseigbe D. A. (1996). Occurrence of Mycollora and Aflatoxins in Marketed Tiger Nut. *Crop Research.* 11: 219 -223.

- 8. Bixquert, M. (2003). Digestive Aspects of Tiger nuts, Tiger nuts Traders. S.L. www.tigernuts.com Bosch, L., Alegrla, A. and Farre, R (2005) RP-HPLC Determination of Tiger Nut and Orgeat Amino Acid Contents: SAGE Publications. Food Science and Technology.11: 33-40.
- 9. Bosch, L. Alegra, A. and Farre, R. (2005). RP-HPLC Determination of Tigernut and Orgeat Amino Acid Contents: SAGE Publications. Food Science Technology. 11: 33-40.
- 10. Deatra, J. and Sams, J.Y. (2009). Pest or Crop and the Future of Southern agriculture. A comprehensive review on its traditional medicinal uses, phytochemicals, pharmacological activities, mechanisms of action and toxicity. Arabian Journal of Chemistry. 18(10): 234-240.
- 11. Delzenne, N.M. (2003).
 Oligosaccharides: State of the Art. Proclamations of the Nutritional Society, 62: 177-182 De Vries, Femke T. 1991. Chufa (Cyperus esculentus, Cyperaceae): A Weedy Cultivar or a Cultivated Weed. Ecology and Botany. 45: 27-37.
- 12.De Vries, Q.S. and Femke, T. (1991). Chufa (Cyperus esculentus, Cyperciceae: A Weedy Cultivar or a Cultivated weed. Journal of Neurochemistry.88: 63-69.
- 13.Eka, O. U. (1998). Roots and Tubers in: Nutrition Quality of Plants Foods Edited by Osagie and Eka. Published by the Post Harvest Research Unit of the Department of Biochemistry University of Benin, Benin City Nigeria. Pp 1-31.
- 14.FAO (1988). Traditional Food Plants: Food and Nutrition

- Paper 42 Rome 239-242.
- 15.FAO / WHO/ UNU (2002). Human, Vitamin and Mineral Requirements (RecommendedDietary Intakes): Report of a Joint Food and Agricultural Organization, World Health Organization and United Nations University. Arabian Journal of Chemistry. 18(10): 234-240.
- 16. Gibson, G. R. (1999). Dietary Modulation of the Human Gut Microflora using the Prebiotics Oligofructose and Inulin. *Journal of Nutrition*. **129**: 143-149.
- 17. D'Onofrio. F. (1997). Larginine for Testing Endothelium-dependent Vascular Functions in Health and Disease. *American Journal of Physiology*. **273**: 606-612.
- 18. Grossman, A. C. and Thomas, L. G. (1998). The Horchata Factory: Origin of the Word Horchata and the Beverage. Journal of Pharmaceutical Biological and Chemical Science.8(5): 305-310.
- 19.IBR (2005). Rhizome. Highbeam research Information. A systematic review of its research history and future prospects. Asian Pacific Journal of Tropical Medicine.10: 835-848.
- 20. Hlilton, E. J. (1976) Catering: Food and Drink Macdonald and Evans Limited. Estover, Plymouth. Pp 152-153.
- 21.Hu, Stampfer, M.J., F.B., Manson, E., J.E., Rimm, Colditz, G.A., Rosner, B.A., Hcnnekens, C.H. and Willett, W.C. (2007). Dietary Fat Intake and the Risk of Coronary Heart Disease in Women. New Enaland Iournal of *Medicine.***20**: 337.

- 22.IHS (2005). In Heat Scent: Chufa. review of Α its traditional uses. isolated biological acetogenins and International activities. of Journal Molecular Science.16: 15625-15658.
- 23. Lapham, J and Drennan, D.S.H. (1990). The Fate of Yellow Nutsedge (Cyperus esculent us): Seed and Seedlings in Soil. Weed Science. 38: 125-128.
- 24. Lopez-Ridaura, R., Willett, W.C., Rimm, E.B., Liu, S., Stampfer, M.J., Manson, I.E. and Hu. F.B. (2004). Magnesium Intake and Risk of Type 2 Diabetes in Men and Women. *Diabetes Care.* **27**: 134-40.
- 25. Negbi, M. (1992). A Sweetmeat Plant, a Perfume Plant and Their Weedy Relatives: A Chapter in the History of Cyperus esculentus L. and C. rotundus L. Ecology Botany. 46: 64-71.
- 26. Nelson, John H., and Kenneth C. Kemp (2006). Laboratory Experiments, Chemistry: TheCentral Science. 10th ed. Upper Saddle River, NJ: Pearson Prentice Hall, pg 87.
- 27.NUTRA (2005). Non-dairy Drinks Easy Pushover for Soy? Food and Beverage and ingredients Development. International Journal of Molecular Science. 19: 156-158.
- 28. Oladele, A.K. and Aina, J.O. (2007). Chemical composition and functional properties of flour produced from two varieties of tigernut. *African Journal of Biotechnology'*.**6**: 2473-2476.
- 29. Pieper, G.M., Jordan, M., Adams, M.B. and Roza, A.M. (2006). Restoration of Vascular Endothelial Function in Diabetes: *Diabetes Research in Clinical Practice*. Pp. 157-

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162.

- 30. Sheen, R.T. and Kahler, H.L. (1999). Effect of Ions on Mohr Method Chloride for Determination. Industrial and Engineering Chemical Analysis. 10(11): 628-629.
- 31.Skoog, D.A.; West, D.M.; Holler, F.J. (2006).Fundamentals of Analytical Chemistry, 7^{th} Edition Thomson Learning, Inc, USA. Pp. 45.
- 32. Temple, V.J. (2008); Nuts and Seeds in: Nutrition Quality of Plants Foods Edited by Osagie and Eka. Published by the Post Harvest Research Unit of the Department of Biochemistry University of Benin, Benin City Nigeria. Pp. 245-274.
- 33. Temple, V.J., Ojobe, T.O. and Kapu, M.M. (2000). Chemical Analysis of Tiger Nut (Cyperus esculentus L). Journal of the of Food Agriculture.50: 261-263.
- 34. Umierie, S.C. and Enebeli, J.N. (1996). Malt Caramel from Tubers of Cyperus esculentus Copyright Elsevier Sci. Ltd Great Britain. Bioresource and Technology.57: 215-219.
- 35. Umerie, S.C. and Uka, A. S. (1998).Brew wort from Cyperus esculenius Tubers. Elsevier *Bioresource* Technology.66: 83-85 100.
- 36. Venho, B., Voutilainen, S., Valkonen, V.P., Virtanen, J., Lakka. T.A., Rissanen. T.IL. Ovaskainen, M.L., Laitinen, M. (2002).and Salonen, J.T. Arginine intake. pressure and the Incidence of Acute Coronary Events in Men: the Kuopio Ischaemic Heart Disease Risk Factor Study. American Journal of Clinical Nutrition.76: 359-364.
- 37. Wu, G., Meininger, C.J., Knabe,

- D.A., Bazer, F.W. and Rhoads, J.M. (2000). Arginine Nutrition in Development. Health and Disease Current Opinion in Clinical Nutrition Metabolic Care.3: 59-66.
- 38. SNWCB (2005). State Noxious Weed Control Board. Yellow Nutsedge (Cyperusesculentus L.). Australian Journal of Basic and applied Science.5(20): 10-30.
- 39.SPSS (2003).Statistical Package for Social Science. Version 12.0.1 for windows.www.spss.com
- 40. Zhang, H. Y., Hanna, M. A., Ali, Y. and Nan, L. (1996). Yellow nut-sedge (Cyperus esculenlus Tuber Oil as a Fuel. Industrial Crops and product. Elsevier. 5: 177-181.