Effect of Ethanol Extract of Anthocleista vogelii leaves on the Liver Parameters of Albino Rats Exposed to Cadmium Oxide

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ABSTRACT

The present study was undertaken to examine the ameliorative potential of ethanol extract of Anthocleista Vogelli leaves on cadmium-induced hepatic damage in albino rats. The liver parameters; Alanine aminotransferase (ALT), Aspartate aminotransferase (AST), Alkaline phosphatase (ALP), Albumin and Bilirubin were examined. Oral administered of cadmium (Cd) (5mg/kg Body weight) to induce toxicity was done once in a week. The extracts (200mg/kg, 400mg/kg and 600mg/kg Body Weight) to the rats were undertaken for 21 days after which liver parameters were measured in the serum using standard methods. The results showed a significant increase in the liver parameters of albino rats exposed to cadmium oxide. Treatments with (200mg/kg, 400mg/kg or 600 mg/kg Body weight) extract in cadmium induce toxicity in albino rats showed a significant (P < 0.05) reduction in the altered biochemical parameters. Hence, the ameliorative effect of Anthocleista vogelii leaves extract against cadmium oxide induced hepatic damage.

Keywords: Anthocleista vogelii, Liver, Cadmium oxide and Hepatotoxicity.

INTRODUCTION

Many scientific studies are being carried out to investigate heavy metals poisoning and toxicity, and how they affect our human existence; particularly our health. Cadmium oxide, a heavy metal compound is one of those heavy metals being studied due to its deleterious effects in the body [1, 2, 3, 4, 5, 6]. Some documented deleterious effect of cadmium toxicity includes; hepatocellular damage. testicular atrophy, hypertension, renal dysfunction, anemia and injury to the central nervous system [7, 8, 9, 10, 11]. In 2010, the World Health Organization (WHO) has published a list of 10 chemicals or groups of chemicals of concern for human health, which includes cadmium [12, 13, 14, 15]. Additionally, the US Agency for Toxic

Substances and Disease Registry (ATSDR) ranked Cadmium in seventh place on the priority list of dangerous substances [16, 17, 18, 19, 20]. Due to its ubiquitous form in nature, cadmium oxide is widely distributed in the atmosphere, water and food, and when exposed to humans in low concentration can cause serious health problems and probably death [21, 22, 23]. Sources of cadmium exposure to humans include fossil fuels, iron and steel production, cement, nonferrous metals production, waste incineration, smoking, fertilizers, etc., and are supplied indirectly to the environment by activities like volcanic eruption, mining and use of phosphate fertilizers [24, 25, 26]. The mechanism of its excretion in humans is not particularly understood, hence it

bioaccumulates in various tissues especially kidneys, lungs, pancreas and liver [27, 28, 29]; causing dysfunction in various animal and human organs notably the kidney, brain, testes, heart and liver [30,31, 33].

In the liver, cadmium is taken into the hepatocytes where it binds to metallothioneins (MTs), glutathione (GSH) and other proteins or peptides and form new complexes. Hydroxyl radical is the most reactive and radical formed; damaging which initiates cellular damages and lipid peroxidation (LPO) even at extremely small concentrations [14, 17, 18, 19], thus the aetiology of hepatotoxicity is established. Hepatotoxicity refers to liver dysfunction or liver damage that is associated with an overload of drugs or xenobiotics [18]. Certain biomarkers used to monitor hepatoxicity include aminotransferase Alanine (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP) and bilirubin [13].

The global burden of hepatotoxicity fifty million affects over people worldwide [20]. Medicinal plants are known to posses antioxidant properties they contain appreciable because minerals. Hence, in order to ameliorate cadmium induced hepatic injury; medicinal plants might be useful due to their relative availability, low cost and

reduced side effects [21]. Various parts of medicinal plants have been used traditionally to mitigate and treat human ailments [25]. Meanwhile, researchers have studied plant extracts and isolated secondary metabolites to establish their pharmacological effects in *in-vivo, ex-vivo*, and *in-vitro* models [4, 7, 9, 10] for the treatment of various ailments.

Anthocleista vogelii is a medicinal plant that is used in treatment of diseases and swelling in the body [11]. It belongs to the family Gentiamuceae, an erect, cylindrical tube of about 20m tall [14]. Traditionally, the leaves and stem-bark are known for treating swellings in the body (anti-inflammatory) [8], while the root-bark and leaves are used in local medicine [9] Its potency as drug is revealed in phytochemical studies of the leaves carried out by [15]. Phytochemicals such as flavonoid, saponins, alkaloid, sterols, phenols, and terpenes were reported to be present. The presence of these secondary metabolites in them has been evaluated different medicinal in plants for treatment of ailments locally, thus making the plant species. a subject of therapeutic interest. This paper seeks to provide a review on the use of Anthocleista vogelii leaves extract in the treatment of cadmium oxide induced hepatotoxicity.

MATERIALS AND METHODS

Plant Collection

The fresh leaves of *Anthocleista vogelii* was collected in a nearby bush beside Presco campus, Ebonyi state University, Abakaliki. The leaves were identified by

a botanist, Prof.E.Onyekweru of Applied Biology Department, Ebonyi State University Abakaliki.

Sample Processing

The sample was allowed to air-dry and turn crispy. The dried leaf sample was ground into power using mechanical grinder and sieved to remove chaff. The fine powders were packed into an airtight container ready for analytical use.

Preparation of Extract

Exactly, 250g of the powdered sample was weighed into 500ml of absolute ethanol in a plastic container. This was allowed to stand for 72hours with intermittent shaking. The mixture was filtered with Whatman No. 1 filter paper under reduced pressure. The resulting filtrate was evaporated using rotary evaporator. Packed in airtight container and stored in refrigerator at 4°c pending further studies.

Preparation of Vitamin C

Vitamin C was obtained from Octovia Pharmarcy, Water Works Road Abakaliki Ebonyi State University. Exactly, 2g (5 tablets) of Vitamin C tablet was dissolved in 100ml of distilled water to obtain stock solution of 0.02g/ml.

Preparation of Cadmium oxide

Exactly 2g of cadmium oxide powder was dissolved in 100ml of distilled water to obtain a stock solution of 0.02g/ml.

Collection of animals

Adult albino rats were sourced from the animal farm of University of Nigeria Nsukka (UNN), weighing 200g-250g.They were kept in well ventilated and hygienic condition in the animal house of Biochemistry Department, Ebonyi State University, Abakaliki, Nigeria. During acclimatization, the animals had free access to animal grower feed and clean water.

Experimental Design

Thirty-six adult albino rats were obtained from the Animal house of the Department of Biochemistry, Faculty of Science, University of Nigeria Nsukka (UNN), weighing between 200 g and 250 g. The animals were allowed access to feed and water for a period of fourteen days, for their acclimatization prior to the commencement of the experiment. The animals were kept in well ventilated cages at room temperature (28° - 30 °C), and under control lights. The rats were randomly distributed into six groups of six animals each. Group 1: served as the normal control and consisted of animals fed with rat pellet and water only. Group 2: consisted of animals fed with rat

After twenty-one (21) days, the rats were sacrificed and blood sample collected

pellet and cadmium oxide only (positive control). Group 3: consisted of animals fed with rat pellet, cadmium oxide and Vitamin C (Standard control). Group 4: consisted of animals fed with rat pellet, cadmium and A. Vogelii extract (200 mg/kg). Group 5: consisted of animals fed with rat pellet, cadmium oxide and A. Vogelii extract (400 mg/kg), while Group 6: consisted of animals fed with rat pellet, cadmium oxide and A. Vogelii extract (600 mg/kg). A. Vogelli extracts (200, 400 and 600 mg/kg bw) was given orally by intubation for 21 days. Cadmium oxide was administered twice in a week (200 mg/l CdO) in the animals for three weeks to induce toxicity.

Collection of blood sample

through venous puncture into a sterile bottle for analysis.

EXPERIMENTAL PROCEDURES

Quantitative estimation of serum alanine transaminase (ALT) and asapartate transaminase (AST): The method of Reitman and Frankel, (1975) was used.

Quantitative estimation of serum alkaline phosphatase (ALP) : Alkaline phosphatase in the serum was estimated by the end point colorimetric method (Englehardt, 1970). **Determination of albumin**: Albumin concentration was determined in serum according to the method of Doumas (1971).

Determination of Total Bilirubin: Total bilirubin was determined according to the method by Schmidt and Eisenburg (1975).



Figure 1. Plot of Aspartate level in different groups (**Group 1:** Control (Nothing given, only feed), **Group 2:** Heavy metal given, **Group 3:** Heavy metal + Vitamin C, **Group 4:** 200mg + Heavy metal, **Group 5:** 400mg + Heavy metal, **Group 6:** 600mg + Heavy metal).



RESULTS

Figure 2. Plot of ALT level in different groups (**Group 1**: Control (Nothing given, only feed), **Group 2**: Heavy metal given, **Group 3**: Heavy metal + Vitamin C, **Group 4**: 200mg + Heavy metal, **Group 5**: 400mg + Heavy metal, **Group 6**: 600mg + Heavy metal).



Figure 3. Plot of ALP level in different groups (**Group 1:** Control (Nothing given, only feed), **Group 2:** Heavy metal given, **Group 3:** Heavy metal + Vitamin C, **Group 4:** 200mg + Heavy metal, **Group 5:** 400mg + Heavy metal, **Group 6:** 600mg + Heavy metal).



Figure 4. Plot of Total Bilirubin level in different groups (**Group 1:** Control (Nothing given, only feed), **Group 2:** Heavy metal given, **Group 3:** Heavy metal + Vitamin C, 51

Group 4: 200mg + Heavy metal, **Group 5:** 400mg+ Heavy metal, **Group 6:** 600mg + Heavy metal).

Figure 5. Plot of Bilirubin level in different groups (**Group 1:** Control (Nothing given, only feed), **Group 2:** Heavy metal given, **Group 3:** Heavy metal + Vitamin C, **Group 4:** 200mg + Heavy metal, **Group 5:** 400mg + Heavy metal, **Group 6:** 600mg + Heavy metal).

DISCUSSION

The therapeutic effect of Anthocleista vogelii leaves extract on hepatotoxicity induced by Cadmium oxide was studied. The present study established that administration of aqueous extract of A. *Vogelli* (200,400 or 600 mg/kg BWsignificantly protect the liver against toxicity exerted by cadmium oxide. The synergetic effects of the bioactive constituents present in the extract may be responsible for the observed results. The results showed а significant therapeutic influence on liver parameters treatment groups in administered with the extract. The leaves of A. vogelli have been widely used by natives in treating swellings and generally as a local medicine [8]. Hepatotoxicity induced by cadmium oxide was studied using the following liver biomarkers; Aspartate aminotransferase (AST), Alanine aminotransferase Alkaline (ALT), phosphatase (ALP), Total bilirubin and Albumin. Significant elevations ($p \le 0.05$)

of AST and ALT levels (Fig. 1 & 2) in rats exposed to Cadmium oxide only were observed when compared with the control group. Upon administration of the leaf extract, a significant reduction $(p \le 0.05)$ in AST and ALT levels were observed in the treatment groups (Group 4, 5 & 6) administered with the extract in a dose dependent manner (200 mg/kg, 400mg/kg and 600 mg/kg dose). Both aminotransferases (AST and ALT) are mainly concentrated in the liver; ALT is localized solely in the cytoplasm, whereas AST is present both in the cytosol and mitochondria of hepatocytes [16]. According to [17], Alanine aminotransferase is the most frequently relied biomarker of hepatotoxicity. It is a liver enzyme that plays an important role in amino acid metabolism and gluconeogenesis. It catalyzes the reductive transfer of an amino group from alanine to αketoglutarate to yield glutamate and pyruvate. Elevated level of this enzyme

is released during liver damage. The estimation of this enzyme is a more specific test for detecting liver abnormalities since it is primarily found in the liver [12, 14]. AST levels however, though useful in detecting hepatocellular necrosis, are considered a less specific biomarker enzyme for hepatocellular injury [16].

Bilirubin is an endogenous anion derived from the regular degradation of haemoglobin from the red blood cells and excreted from the liver in the bile. It is a chemical normally present in the blood in small amounts and used by the liver to produce bile. When the liver cells are damaged, they may not be able to excrete bilirubin in the normal way, causing a build-up of bilirubin in the blood and extracellular (outside the cells) fluid. Serum bilirubin could be elevated if its levels build up in blood circulation. Increased levels of bilirubin in the blood may occur due to hepatic damage and can lead to jaundice and other hepatotoxicity symptoms [15]. A significant (p≤0.05) increase in total bilirubin levels were recorded in rats exposed to Cadmium oxide only. The increased serum level of total bilirubin in the present study is a clear indication hepatic dysfunction but, of upon administration of A. Vogelli extract a significant reversal effect was recorded in group 4, 5 and 6. This was in agreement with the result from similar studies carried out by [18, 19]. In acute human hepatic injury, total bilirubin measured is seen as a better indicator of

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disease severity compared to ALT [21]. The ability of A. Vogelli extracts to ameliorate the elevated levels of total bilirubin, also confirms its protective role. High levels of serum alkaline phosphatase (ALP) are also linked to the condition and function of liver cells. An elevated serum alkaline phosphatase is with liver associated damage (Renugadevi and Prabu, 2010). Administration of aqueous extract of A. (200 mg/kg)Voaelli 400 mg/kg and 600mg/kg bw) mitigated cadmiuminduced hepatotoxicity as revealed by the decreased levels of ALP, AST and ALT compared with the cadmium oxide exposed in group: G2. [16] reports that ALP plays an important role in deciding the type of liver damage bv which hepatotoxins. Albumin, is manufactured by the liver, is a major protein that circulates in the blood stream. Albumin concentration is a biochemical parameter for monitoring liver function in the blood [16]. Abnormal levels of this protein have been reported to be associated with liver damage [20] Albumin concentration in the serum of albino rats exposed to cadmium oxide were significantly lower (p<0.05) in comparison to the treatment groups. In the present study, the significant decrease in the serum albumin of animals exposed to cadmium oxide can be attributed to impairment in hepatocyte functions causing decreased cvtochrome P-450 activity and inhibition in protein metabolism in the liver [15].

CONCLUSION

The outcome of this study clearly indicates that the ethanol extract of *Anthocleista vogelii* leaves ameliorated the altered liver function parameters triggered by cadmium oxide-induced hepatotoxicity in the biochemical parameters. The plant therefore could serve as a possible potent herbal drug for the correction of liver malfunction.

REFERENCES

 Abdel Moneim, A. E., Bauomy, A. A., Diab, M. M., Shata, M. T., Al-Olayan, E. M. and El-Khadragy, M. F. (2014). The Protective Effect of Physalis Peruviana L. Against Cadmium-Induced Neurotoxicity in Rats. *Biological Trace Element Research*; **160**: 392-99.

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- Aggarwal, B. B., Prasad, S. and Kannappan, R, (2011). Identification of Novel Anti-Inflammatory Agents from Ayurvedic Medicine for Prevention of Chronic Diseases: "Reverse Pharmacology" and "Bedside to Bench" Approach. *Curr Drug Targets*; 12:1595-1653.
- 3. Asagba, S. (2010). Alteration in the activity of oxidative enzymes in the tissues of male wistar albino rats exposed to cadmium. *International Journal of Occupational Medicine and Environmental Health*, 23: 55-62.
- 4. Atanasov, A., Waltenberger, B. and Pferschy-Wenzig, E. (2015). Discovery and Resupply of Pharmacologically Active Plantderived Natural Products: A Review. *Biotechnology Advancement.*; **33**:1582-1614.
- 5. ATSDR Substance Priority List |ATSDR. Available online: https://www.atsdr.cdc.gov/spl/
- Babatunji, E. O., Basiru, O. A., Oluwafemi, A. O., Habiba, M. M.Sunday, A. O. and Abiodun, A. O. (2016). Ameliorative potential of *Aframomum melegueta* extract in cadmium-induced hepatic damage and oxidative stress in male Wistar rats. *Journal of Applied Pharmaceutical Science;* 6(07): 094-099.
- Amacher, D. E. (2002). A Toxicologist's Guide to Biomarkers of Hepatic Response. *Human Experimental Toxicology;* 21: 253-262.
- Bruha, R., Dvorak, K. and Petrty, J. (2012). Alcoholic Liver Disease. *World Journal of Hepatology*; 4:81-90.
- 9. Dufour, D. R., Lott, J. A., Nolte, F. S., Gretch, D. R. and Koff, R. S. (2000). Diagnosis and Monitoring of Hepatic Injury: I. Performance

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- 10. Dufour, D. R., Lott, J. A., Nolte, F. S., Gretch, D. R. and Koff, R. S. (2001). Diagnosis and
- 11. Monitoring of Hepatic Injury. II. Recommendations for Use of Laboratory Tests in Screening, Diagnosis and Monitoring. *Clinical Chemistry;* **47**: 1133-1135.
- 12. Doumas, B. T., Watson, W. A. and Biggs, H. G. (1971). Determination of Serum Albumin. *Clinical Chemistry Acta.*; 31:87-89.
- 13. Elgaml, S.A. and Hashish, E.A. (2014). Clinicopathological studies of *Thymus vulgaris* Extract Against Cadmium Induced Hepatotoxicity in Albino Rats. *Global Journal of Pharmacology*, 8: 501-09.
- 14. Englehardt, V.A. (1970). Measurement of Alkaline Phosphatase. *Aerzt Labour;* **16**:42
- 15. Gambini, J., Inglés, M. and Olaso, G. (2015). Properties of Resveratrol: in vitro and in vivo studies about Metabolism, Bioavailability, and Biological Effects in Animal Models and Humans. Oxid MedCell Longev.; 837042.
- 16. Honey, S., Neetu, R. and Blessy,
 B. M. (2015). The Characteristics, Toxicity and Effects of Cadmium. *International Journal of Nanotechnology and Nanoscience*, 3: 1-9
- 17. Haidry, M. T. and Malik A. (2014). Hepatoprotective and Antioxidative Effects of Terminalia Arjunaagainst Cadmium Provoked Toxicity in Albino Rats (Ratus Norvigicus). *Biochemical Pharmacology*; 3: 130.
- 18. Jeyaprakash, K. and Chinnaswamy, P. (2005) Effect of

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Spirulina and Liv-52 on cadmium induced toxicity in albino rats. *Indian Journal of Experimental Biology* 43(9):773

- Mitra, E., Ghosh, A. K., Ghosh, D., Mukherjee, D., Chattopadhyay, A., Dutta, S., Pattari, S. K. and Bandyopadhyay, D. (2012). Protective Effect of Aqueous Curry Leaf (Murraya Koenigii) Extract against Cadmium-Induced Oxidative Stress in Rat Heart. Food and Chemical Toxicology.; 50: 1340-53.
- 20. Meagan, T., Yogini, J., Ilya, W. and Leonard, W. (2017). Hepatotoxicity: Treatment, Causes and Applications of Medicinal Plants as Therapeutic Agents. *The Journal of Phytopharmacology*, **6**(3): 186-193
- 21. Nair, A. R., DeGheselle, O., Smeets, K., Van Kerkhove, E. and Cuypers, A. (2013). Cadmium-Induced Pathologies: Where is the Oxidative balance Lost (or not)? *International Journal of Molecular Sci*ences; 14: 6116-43.
- 22. Navarro, V. J. and Senior, J. R. (2006). Drug- Related Hepatotoxicity. *New England Journal of Medicine.*; **354**:731-739.
- 23. Ozer, J., Ratner, M., Shaw, M., Bailey, W. and Schomaker, S. (2008). The Current State of Serum Biomarkers of Hepatotoxicity. *Toxicology*; 245: 194-205.
- 24. Page, A. L., and Bingham, F. T. (1973). Cadmium residues in the environment. *Residue Reviews*, pp.1-44).
- 25. Reitman S, and Frankel S (1957). A colorimetric method for the determination of serum glutamateoxaloacetate and pyruvatetransaminase. *American Journal of Clinical Patholology*. 28: 56-63.

26. Renugadevi, J.and Prabu, S.M. (2010) Cadmium-induced hepatotoxicity in rats and the protective effect of naringenin. *Experimental Toxicological Pathology* 62(2):171–181

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- 27. Saukkonen, J. J., Cohn, D. L., Jasmer, R. M., Schenker, S. and Jereb, J. A. (2006). An Official ATS Statement: Hepatotoxicity of Antituberculosis Therapy. *American journal of Respiratory and Critical Care Medicine.;* **174**: 935-952.
- 28. Sarkar, A., Ravindran, G. and Krishnamurthy, V. (2013). A brief Review on the Effect of Cadmium Toxicity: From Cellular to Organ Level. International Journal of Biotechnology Research,; **3**:17-36.
- 29. Singh, A., Bhat, T. K. and Sharma, O. P. (2011). Clinical Biochemistry of Hepatotoxicity. *Journal of Clinical Toxicology*, **S4**:001.
- 30. Schmidt, M. and Eisenburg, J. (1975). Serum Bilirubin Determination inNewborn Infants. A New Micro-method for the Determination of Serum of Plasma Bilirubin in Newborn Infants. Fortschr Med.; 93:1461-66.
- 31. Vicente-Sánchez, C., Egido, J., Sánchez-González, P.D., Pérez-Barriocanal, F., López-Novoa, J.M. and Morales, A.I. (2008) Effect of the flavonoid quercetin on cadmium-induced hepatotoxicity. *Food Chemistry Toxicology* 46(6):2279-2287
- 32. WHO (2010). Action Is Needed on Chemicals of Major Public Healt Concern. *Public Health Environmental*, pp. 1-4.
- 33. Zhang, T., Hu, Y., Tang, M., Kong, L., Ying, S.J., Wu, T., Xue, Y. and Pu, Y. (2015). Liver toxicity of cadmium telluride quantum dots (CdTe QDs) due to oxidative stress in vitro and in vivo.

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www.idosr.org International Journal of Molecular Science 16: 23279-299.

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